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(54) MÉTHODE ET DISPOSITIF POUR DOSER DES SUCCÉDANES
DU SANG

(54) METHOD AND APPARATUS FOR MEASUREMENT OF BLOOD
SUBSTITUTES

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(57) L'invention concerne une méthode qui permet d'identifier et de quantifier rapidement et précisément la concentration d'un succédané de sang, tel que l'hémoglobine réticulée, dans un échantillon de sérum ou de plasma. Selon la méthode, on mesure la concentration du succédané de sang et on utilise cette mesure pour corriger son effet, s'il existe, sur la concentration mesurée d'un analyte, tel que les protéines totales du sérum/plasma. La méthode permet de déterminer la concentration de l'hémoglobine vraie en présence des succédanés de sang. Elle est appliquée aux échantillons contenus dans un tube étiqueté primaire ou secondaire, ou à l'extrémité d'une pipette utilisée pour dispenser le sérum ou le plasma dans un analyseur.

(57) A method is disclosed whereby the concentration of a blood substitute, such as cross-linked hemoglobin, in a serum or plasma specimen is rapidly and accurately identified and quantified. The method further takes the measured concentration of the blood substitute and uses it to correct for its effect, if any, on a measured analyte concentration, e.g., serum/plasma total protein. Further, the method allows for the determination of the concentration of true hemoglobin in the presence of blood substitutes. The method is carried out in respect of samples contained in a primary or secondary labelled tube, or a pipette tip used to dispense serum or plasma in a blood analyzer.





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(54) Title: METHOD AND APPARATUS FOR MEASUREMENT OF BLOOD SUBSTITUTES

(57) Abstract

A method is disclosed whereby the concentration of a blood substitute, such as cross-linked hemoglobin, in a serum or plasma specimen is rapidly and accurately identified and quantified. The method further takes the measured concentration of the blood substitute and uses it to correct for its effect, if any, on a measured analyte concentration, e.g., serum/plasma total protein. Further, the method allows for the determination of the concentration of true hemoglobin in the presence of blood substitutes. The method is carried out in respect of samples contained in a primary or secondary labelled tube, or a pipette tip used to dispense serum or plasma in a blood analyzer.

WE CLAIM:

1. A method of identifying the presence of a selected interferent contained in a specimen in the presence of one or more other interferents, by quantification of the selected interferent in said specimen, using a spectrophotometer, comprising the steps of:
 - (i) generating a calibration algorithm for said selected interferent;
 - (ii) measuring with said spectrophotometer, absorbance of radiation by said selected interferent in said specimen; and
 - (iii) incorporating said absorbance measured in step (ii) in said algorithm and calculating the concentration of said selected interferent in said specimen.
2. The method of claim 1 wherein said selected interferent is a blood substitute and said one or more other interferents are selected from the group consisting of haemoglobin liberated from blood cells, turbidity and bile pigments.
3. The method as claimed in claim 1 wherein said selected interferent is CLHb and said one or more other interferents are haemoglobin liberated from blood cells, turbidity and bile pigments, and said algorithm is:
$$g/L \text{ CLHb} = A(541\text{nm}) - B(558\text{nm}) + C(600\text{nm}) - D(616\text{nm}) + E$$
where (X_{nm}) is the first derivative of absorbance measured at the wavelength specified and A, B, C, D and E represent constants.
4. The method of claim 1 wherein said selected interferent is haemoglobin liberated from blood cells, and said one or more other interferents is a blood substitute.
5. The method of claim 4 wherein said concentration of

liberated Hb is determined in the presence of one or more additional interferents chosen from the group consisting of intralipid (IL), bilirubin (BR) and biliverdin (BV).

6. The method of claim 5 wherein said calibration algorithm for liberated Hb is:

$$\text{g/L Hb} = A(558) - B(570) + C(730) - D$$

where numbers in the parenthesis are the first derivative of absorbance at the wavelengths (nm) shown and A, B, C and D are constants.

7. The method of claim 1, wherein said quantification includes calculating the first derivatives of at least two portions of a spectrum generated from a scan for a particular interferent which are used to calculate said selected interferent concentration.

8. A method of taking into account the concentration of a blood substitute interferent contained in a specimen, in a measured analyte concentration obtained from a specimen, using a spectrophotometer, comprising the steps of:

- (i) generating a calibration algorithm for said blood substitute interferent;
- (ii) generating an algorithm for said analyte which provides a relationship between a measured analyte concentration and an amount of blood substitute interferent present in said specimen;
- (iii) measuring with said spectrophotometer, absorbance of radiation for said specimen with any blood substitute present in the specimen;
- (iv) using said calibration algorithm and absorbance measured in step (iii) to predict concentration of blood substitute interferent present;
- (v) correlating the relationship from step (ii) and the prediction

from step (iv) to predict concentration of analyte as if no blood substitute interferent were present.

9. The method of claim 8 wherein said specimen also contains one or more non-blood substitute interferents and wherein the concentration of said non-blood substitute interferent is also determined by:

- (i) generating a calibration algorithm for each of at least one non-blood substitute interferent;
- (ii) measuring the absorbance of radiation by said specimen; and
- (iii) correlating the absorbance measured in step (ii) to the amount of said non-blood substitute interferent.

10. The method of claim 8 where the at least one analyte is chosen from the group consisting of Na, K, Cl, HCO₃, Ca, Mg, creatinine, urea, total protein, gamma glutamyl transfurase (GGT), aspartate amino transfurase (AST), lactate dehydrogenase (LDH), creatine kinase (CK), alkaline phosphatase (ALP) and total bilirubin (Tbili).

11. The method of claim 8 where reflectance is used instead of absorbance.

12. The method of claim 8 where the radiation is in the range of 474-910 nm.

13. The method of claim 8 where calibration is conducted with samples containing all interferents expected during an analysis of an unknown sample.

14. The method of claim 12 where the sample contains an even distribution of interferents of interest, and the concentrations of any two interferents do not correlate significantly.

15. The method of claim 9 where said non-blood substitute interferent is selected from the group consisting of haemoglobin (Hb), bilirubin (BR), biliverdin (BV) and turbidity.

16. The method of claim 8 where the measured concentration of an analyte is correlated to an amount of blood substitute present by developing an appropriate linear regression equation in an analyzer.

17. A method of distinguishing true hemolysis from pseudo hemolysis caused by a blood substitute interferent comprising the steps of:

a. identifying the presence of a blood substitute interferent contained in a specimen in the presence of Hb liberated from blood cells, turbidity and bile pigments, by quantification of the blood substitute in said specimen, using a spectrophotometer, comprising the steps of:

- (i) measuring absorbance of radiation by said interferent in said specimen; and
- (ii) incorporating said absorbance measured in step a(i) in the following algorithm:
$$g/L \text{ CLHb} = A(541\text{nm}) - B(558\text{nm}) + C(600\text{nm}) - D(616\text{nm}) + E$$

where (X_{nm}) is the first derivative of absorbance measured at the wavelengths specified and A, B, C, D and E represent constants; and calculating the concentration of said interferent in said specimen; and

b. measuring Hb liberated from blood cells in the

presence of said blood substitute interferent contained in a specimen, using a spectrophotometer, comprising the steps of:

- (i) measuring absorbance of radiation by said specimen with said blood substitute interferent present; and
- (ii) incorporating said absorbance measured in step b(i) in the following algorithm:
$$g/L Hb = A(558) - B(570) + C(730) - D$$

where numbers in the parenthesis are the first derivative of absorbance at the wavelengths (nm) shown, where A, B, C, and D are constants, and calculating the concentration said liberated Hb in said specimen.

18. The method of claim 17 wherein said concentration of liberated Hb is determined in the presence of one or more additional interferents chosen from the group consisting of intralipid (IL), bilirubin (BR) and biliverdin (BV).

Method and Apparatus for Measurement of Blood Substitutes

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Field of Invention

This invention relates to spectrophotometry and the spectrophotometric analysis of serum or plasma used for blood testing. It also relates to the effects of blood substitutes and other interferents on blood test results. In particular, this invention relates to a method for measuring blood substitutes in serum or plasma, determining the effects of blood substitutes on blood test results, and combining the two to adjust or correct blood test results for the presence of blood substitutes.

15

Background of Invention

Clinical laboratory tests are routinely performed on the serum or plasma of whole blood. In a routine assay, red blood cells are separated from plasma by centrifugation, or red blood cells and various plasma proteins are separated from serum by clotting prior to centrifugation.

Haemoglobin (Hb), bilirubin (BR), biliverdin (BV) and light-scattering substances like lipid particles are typical substances which will interfere with, and affect spectrophotometric and other blood analytical measurements. Such substances are referred to as interferents. Elevated Hb in the blood, haemoglobinemia, can be due to disease states and as a result of specimen handling. Elevated bile pigments, namely BR and BV, can be due to disease states. Increased lipid particles in the blood, also known as hyperlipidemia or lipemia, can be due to disease states and dietary conditions. Lipemia is

the major cause of turbidity in serum and plasma, and therefore the terms lipemia and turbidity are frequently used interchangeably.

Many tests conducted on plasma or serum samples employ a series of reactions which terminate after the generation of chromophores which 5 facilitate detection by spectrophotometric measurements at one or two wavelengths. Measurement of interfering substances prior to conducting such tests is important in providing meaningful and accurate test results. In fact if a sample is sufficiently contaminated with interferents, tests are normally not conducted as the results will not be reliable.

10 Another set of potential interferents is blood substitutes. Blood substitutes are new products which are under development, for use instead of whole blood or red blood cells for transfusion. Blood transfusion is a life-saving process performed after severe blood loss during trauma or surgery. Some advantages of using a blood substitute instead of blood or red blood 15 cells are as follows: 1. blood substitutes are expected to be universally compatible with all blood types, therefore cross-matching will not be necessary; 2. maximum storage time of blood is 42 days, whereas blood substitutes could have a much longer shelf-life; 3. purification a blood substitute may include heat treatment, which may eliminate the threat of hazardous viruses such as HIV. However, a challenge blood substitutes will 20 pose to the clinical laboratory is managing the effects of blood substitutes on blood tests. Some blood substitutes will cause serum or plasma specimens to appear as whole blood or severely hemolyzed serum or plasma, hence the term pseudohemolysis may be used to describe such specimens.

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Hemolysis, the liberation of Hb from red blood cells into serum or plasma, may account for about 2 g/L, but blood substitutes can account for as much as 30 g/L of cross-linked Hb (CLHb), in a patient treated for severe blood loss. However, true hemolysis will not only make serum and plasma specimens appear red, but high concentrations of certain analytes inside red cells, e.g. potassium, will elevate the concentration of analytes in a serum or plasma specimen. Therefore, the effect of Hb-based blood substitutes on blood test results, is more predictable than the effect of true hemolysis.

Current methods used for detecting haemoglobinemia, bilirubinemia and lipemia or turbidity utilize visual inspection of the specimen with or without comparison to a coloured chart.

Most blood substitutes under development are made from human Hb, but another type of blood substitute has been reported which is a milky-white emulsion containing tiny beads of perfluorocarbons wrapped in a surfactant. For example, such perfluorocarbons are described in R.G. Pratt et al. "Quantitation of perfluorocarbon blood substitutes in tissues using F-19 magnetic resonance spectroscopy" Biomaterials, Artificial Cells and Immobilization Technology, Vol. 20, No. 2-4, 1992, pages 921-924, XP002057067. The former will create pseudohemolysis while the latter will create pseudolipemia, in serum and plasma specimens. Subunits of Hb-based blood substitute are chemically cross-linked for stability (CLHb) and produce absorbance spectra which are very similar to the absorbance spectra of

normal hemoglobin (Hb). A method and apparatus for determining, by absorption spectrophotometry, an analyte, such as bilirubin or hemoglobin, in human serum, in the presence of unknown interferents is described in European Patent Application 0 158 506. However, the method relies upon wet chemistry and dry chemistry assays in order to make the analyte determinations.

As such, currently there is no method for rapidly adjusting blood test results which are effected by blood substitutes. The present Invention describes such a method. The method and apparatus of the present invention for measuring the concentration of blood substitutes in the presence of Hb, BR, BV and turbidity: The results of measurements of blood substitutes obtained

are used to correct blood test results affected by the blood substitutes.

Further the levels of other interferents can be measured in the presence of the blood substitutes. Therefore, true hemolysis can be measured in the presence of blood substitutes which cause "pseudo-hemolysis".

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Summary of the Invention

According to one aspect of the present invention there is provided a method whereby the concentration of a blood substitute in a serum or plasma specimen is rapidly and accurately assessed, and the measured concentration is used to correct for its effect, if any, on a measured analyte concentration, e.g., serum/plasma total protein. There is no known prior art which can measure this interferent in a primary or secondary labelled tube, or a pipette tip used to dispense serum or plasma in a blood analyzer.

According to a further aspect of the present invention, spectral data is used in a novel way to determine if a specimen contains a blood substitute and if so, to what extent, in the presence and absence of other interferents.

According to another aspect of the present invention, other interferents, for example Hb, BR, BV and turbidity, can be accurately measured in the presence of blood substitutes.

According to still another aspect of the present invention, linear regression equations may be developed for different analytes in different blood analyzers by which it is possible to relate the measured concentration of an analyte to the amount of blood substitute present. The analyzers examined are the Kodak Ektachem 700 from Johnson and Johnson and the

Hitachi 717 from Boehringer Mannheim; the former represents slide technology or the so-called dry chemistry (i.e., the only liquid involved in the test is the specimen being tested), and the latter represents the so-called wet chemistry. The blood substitute tested was CLHb from Hemosol Inc.

5 According to yet another aspect of the present invention, algorithms are developed which are analyte-specific and analyzer-specific, which can measure the amount of blood substitute present in the serum or plasma specimen, and make the necessary adjustment or correction to the measured analyte concentration, thereby removing the effect of the blood substitute.

10 Other advantages and novel features of the invention will become more apparent from the following detailed description when taken in conjunction with the accompanying drawings.

Description of the Drawings

15 **Figure 1** is a graphic representation of a linear regression fit of the data generated from the CLHb calibration.

Figure 2 provides a graphic representation of the results of linear regression fit for predicted CLHb concentration, for samples not used in the calibration processes.

20 **Figure 3** provides a representation of the results of a linear regression fit of data generated from true Hb calibration in the presence of cross-linked Hb and other interferents (IL, BR, BV).

Figure 4 is a graphical representation of a linear regression fit for data in respect of total protein.

Figure 5 is a graphical representation of linear regression fit for data in respect of aspartate aminotransferase.

Figure 6 is a graphical representation of linear regression fit for data in respect of alkaline phosphatase.

5 **Figure 7** is a graphical representation of linear regression fit for data in respect of total bilirubin.

Definitions

Abbreviations as used herein have the following meaning:

10	Na	=	sodium
	K	=	potassium
	Cl	=	chlorine
	HCO ₃	=	bicarbonate
	Ca	=	calcium
15	Mg	=	magnesium
	GGT	=	gamma glutamyl transfurase
	AST	=	aspartate amino transfurase
	LDH	=	lactate dehydrogenase
	CK	=	creatine kinase
20	ALP	=	alkaline phosphatase
	Tbili	=	total bilirubin

Detailed Description of the Invention

The apparatus used in the present invention generally comprises a dual-beam-in-time spectrophotometer optically coupled to, or communicating with a sample holder through optical fibres. The sample can be a serum or plasma specimen in a labelled tube made of glass or plastic material or a plastic pipette tip used for dispensing the specimen in the blood analyzer. A robotic arm can be used to introduce the sample into the sample holder. It is understood that other conveyor transport mechanisms could be employed. Furthermore, any means by which the input and output fibre optic bundles are brought into alignment for measurement of absorption or reflectance in a specimen container is within the scope of the present invention. Along the side of the sample holder is a separate fibre for transmitting the reference light, when the shutter at the sample channel is closed and the shutter at the reference channel is open. Sample and reference dark scans can be performed with the sample in place with the sample holder closed, and both shutters closed, using the integration times used for the respective sample and reference light scans. There are no other shutters in the apparatus besides the sample path and reference path shutters. The spectrophotometer uses a fixed integration time for the reference beam and a choice of integration time for the sample beam.

Preferably the light source is a quartz-tungsten-halogen 10 watt lamp, but other wattage lamps can be employed. The input power supply is alternating current, but the output to the light source is a stabilized direct current. The spectral output from the light source is broad band covering the visible and NIR regions. The beam of radiation from light source is directed

through a band-pass filter in the lamp assembly, and a shaping filter in the spectrophotometer. The band-pass filter is required to reduce unwanted radiation outside of 475-910nm, and the shaping filter is required to "flatten" the detection system's optical response. The 475-910nm wavelength range is used because this range can be used to measure the blood substitutes as well as other interferences, i.e., Hb, BR, BV and turbidity. A bifurcated bundle emerges from a main fibre connected to the output light source which provides random sampling of the lamp radiation to supply the sample and reference beams via two arms. The sample and reference beams reconverge at the spectrophotometer via the two arms of another bifurcated fibre bundle. In order to provide balanced emerging radiation to the linear photodiode array (PDA) detector, from both the sample and reference paths, 99% and 1% of the radiation from the main fibre must be directed through the sample and reference paths, respectively.

If the sample holder is not light-tight, i.e., capable of excluding external light, sample and reference dark scans have to be subtracted from the sample and reference light scans, respectively. If the sample holder is sufficiently light-tight, which is determined empirically, dark scans are omitted. Also, multiple scans can be averaged to minimize noise, but for the sake of speed single scans are used.

When in use, each pixel or wavelength portion is measured at the photodiode array approximately simultaneously during a particular scan. The optical radiation falling on each sensor element is integrated for a specified

time and the individual pixels or wavelengths are sampled sequentially by a 16 bit analog-to-digital convertor or ADC.

Although the present embodiment details use of a PDA, any alternative means which provides a similar result is understood to be within the scope of the present invention. For example a filter-wheel system may be used. In carrying out measurements each analyte uses from one to four wavelengths or pixels. Given that the first derivative of absorbance with respect to measurements with the PDA is the difference between the absorbance at two adjacent pixels, the first derivative of absorbance at one wavelength with a filter-wheel system will require the absorbance measured with two different narrow band-pass filters. It will be readily understood by those skilled in the art that the filters do not need to be assembled on a rotating wheel, but that any structure which achieves the result of a narrow band-pass filtration of absorbed radiation is within the scope of the present invention.

Another embodiment uses reflectance where the incident light is reflected off some reflective surface placed behind the sample.

Transmission is preferred over reflectance because the effect of the variation in absorbance due to the container in which the serum or plasma specimens reside, are insignificant for the wavelengths chosen. The variation in apparent absorbance due to markings on the labels can be handled effectively by using the first derivative of the apparent absorbance. The term "apparent" absorbance is used because when the amount of light transmitted through the sample is measured, and the transmitted light is

converted to absorbance units, as discussed in the next paragraph; light attenuation by any means other than that which is absorbed by the sample will be interpreted as absorbance. For example, lipid particles will scatter light away from the detector, and the scattered light will be interpreted as absorbance. It is understood that determination of the concentration of interferents using reflectance is within the scope of this invention.

The PDA integrates the optical radiation over a specified time and converts the optical signal to a time multiplexed analog electronic signal called a scan where absorbance is calculated as:

$$10 \quad \text{Absorbance}_i = \log\{\text{Reference Light}_i - \text{Reference Dark}_i\} / (\text{Sample Light}_i - \text{Sample Dark}_i) + \log(\text{ITS} / \text{ITR})$$

where $\text{Absorbance}_i = \text{Absorbance pixel } i$
 15 $\text{Reference Light}_i = \text{Reference pixel } i$ readings, with reference path open and sample path closed by a shutter;
 $\text{Reference Dark}_i = \text{Reference pixel } i$ readings, with reference and sample paths closed by shutters;
 $\text{Sample Light}_i = \text{Sample pixel } i$ readings, with sample path open and reference path closed by a shutter;
 20 $\text{Sample Dark}_i = \text{Sample pixel } i$ readings, with sample and reference paths closed by shutters;

ITS = Integration time for sample measurement;

ITR = Integration time for reference measurement;

and

25 $i = \text{the particular pixel (wavelength) in the PDA.}$

Depending on the required analyzer through-put and if the sample holder is sufficiently light-tight, dark scans are not necessary. The electronic signal is proportional to the time that the detector integrates the optical signal. The 30 electronic signal is amplified by analog electronic amplifiers and converted to a digital signal by an analog-to-digital converter or ADC. The digital information from the converter is interpreted for data analysis by a microprocessor which is in turn connected via an RS232 connector to a

computer. The results of the data analysis can be displayed on the computer, or on a printer connected to the computer. A user can control the device through the computer, to specify a particular interferent to be analyzed and to determine the number and timing of measurements.

5 Although a rapid pre-screening device could take as much time as one to two minutes per sample measurement and still be considered rapid in this field of art, the present invention allows for rapid pre-screening of samples by taking successive sample measurements at intervals of 5 seconds for 4 interferents and the blood substitute; for pipette tips and if no dark scan
10 measurements are made with same tips, the spectral measurements can be made in up to one second. After the sample holder is opened, the sample is placed according to the controlling process and a sensor in the sample holder can activate the collection of spectral data.

15 The integration time for the sample beam is low for clear samples since there is less scattered light and therefore more light is transmitted to the detector. When the light is sufficiently scattered by, for example a turbid sample, the spectrophotometer will automatically switch to a higher integration time. Due to the linear relationship between the detector response and the IT, the second IT can be determined from the detector
20 response of the low IT (or first IT). The higher integration time chosen will be within a pre-selected range such that the detector's response is optimal and within the linear-response range. This feature will allow all samples, from the clearest to the most turbid, to be efficiently screened without exceeding the linear response range of the detector.

As with any quantitative method, calibration of the spectrophotometer is required. However the method for NIR calibration is much more complex than most which can be calibrated with a minimum of a single standard material of known concentration. In respect of NIR calibration, samples must contain all interferents expected during the analysis of an unknown sample; the sample must contain an even distribution of the interferent of interest, and the concentrations of any two interferents should not correlate significantly. It is understood, that for any pre-screening, according to the present invention, of a typical sample for subsequent analysis, any combination of interferents may be present. The pre-screen allows for the determination of the concentration of any one in the presence or absence of the others. Although the data presented is based upon a particular blood substitute made by chemically cross linking the Hb subunits, it is understood that similar calibration algorithms for measuring other blood substitutes and correcting for their effects on blood test results, are within the scope of this invention. It is also understood that not only does "other blood substitutes" include those which are Hb-based, but any other blood substitute, e.g., those which appear as milky-white emulsion.

The first part of the process for generating a calibration curve is to store spectral data for the calibration set. The calibration algorithm for each interferent must be installed in a microprocessor so that when an unknown sample is tested for a particular interferent the result is produced quickly. In order to calculate the quantity of any interferent present, any one of several

different methods, all of which are within the scope of this invention, may be used.

One approach would be to process the raw absorbance by multiple linear regression and choose wavelengths using standard procedures and statistics to find optimal wavelengths at which to describe concentrations of interferences. However significant changes in the spectrum brought about by lipemia, influence the outcome of calculations for haemoglobin or for bilirubin, or biliverdin, or blood substitutes and consequently it is necessary to select additional wavelengths to compensate for these interactions. Nevertheless, 10 this is not a preferred approach.

Another method which may be employed is to use all of the absorbance spectrum, and perform either a principal component analysis or partial least squares analysis and effectively determine from the components that are optimised, the concentration of these different elements. However, 15 a disadvantage of using either of these methods is that they are computationally intensive and consequently take more time to calculate and increase the length of time required to assess each sample.

A preferred method is to calculate the first derivative of certain portions of the absorbance spectra in respect of the particular interferent being measured. It is also possible to calculate the second, or third derivatives of absorbance, and such calculations are within the scope of this invention. However, each step of taking differences to calculate those derivatives is more time consuming and introduces more noise. 20

Smoothing of the absorbence data may be performed if necessary before processing the data.

In practice, an optimal combination of first derivatives of at least two portions of an absorbance spectrum generated from a scan of a plasma specimen containing a particular interferent, is used to calculate interferent concentration. The precise approach used depends on the interferent being measured.

There are three parts to the present invention. The first part of the present invention is to measure the amount of the blood substitute present in the serum/plasma specimen. In respect of Hemosol CLHb optimal results may be obtained by calculating the first derivative of absorbance measurements at wavelengths of approximately 541nm, 558nm, 600nm and 616nm.

The calibration equations outlined below cover the broad range of variability anticipated for the interferents. If low-end accuracy becomes a concern, separate calibrations can be developed: one for the high end, and a second, if the result predicted by the previous calibration is less than a predetermined level.

In order to calibrate the spectrophotometer for CLHb, serum specimens with normal appearance were spiked with 0 to 16.6 g/L CLHb, 0 to 3.2 g/L Hb, 0 to 4.0 g/L IL, 0 to 48.4 mg/dL BR and 0 to 4.0 mg/dL BV as shown in Table 1. No significant intercorrelation among the analytes was allowed. The specimens were run once immediately after preparation, and then repeated using different polypropylene pipette disposable tips. The Hb

was prepared by replacing the normal plasma (by appearance) with water and lysing the erythrocytes through three freeze-thaw cycles. The Hb content of the supernatant of the lysate was measured on an Abbott Cell Dyn.TM The spectra were stored on diskettes. The analyses on a sample 5 sets were performed by a statistical computer program and an algorithm was developed for CLHb. An independent sample sets were set aside for validation (referred to in the graphical representations as prediction) of the calibration equation, and is shown in Table 2.

Sample #	Actual Hemolink (g/L)	Hb (g/L)	IL (g/L)	BV (mg/dL)	BR (mg/dL)	Fitted Hemolink (g/L)
1	4.91	0	0	0	0	4.95
2	4.91	0	0	0	0	5.00
3	2.62	0	0	0	0	2.86
4	2.62	0	0	0	0	2.88
5	0	0	0	0	0	0.19
6	0	0	0	0	0	0.10
7	7.43	0	0	0	0	6.63
8	7.43	0	0	0	0	6.62
9	1.33	0	0	0	0	1.17
10	1.33	0	0	0	0	1.09
11	5.89	0	0	0	0	5.42
12	5.89	0	0	0	0	5.39
13	4.17	2.36	0	0	0	4.51
14	4.17	2.36	0	0	0	4.52
15	7.36	0.77	0	0	0	6.36
16	7.36	0.77	0	0	0	6.54
17	6.58	1.16	0	0	0	6.06
18	6.58	1.16	0	0	0	6.06
19	4.21	1.59	0	0	0	4.51
20	4.21	1.59	0	0	0	4.41
21	1.72	2.03	0	0	0	1.93
22	1.72	2.03	0	0	0	1.92
23	3.33	3.15	0	0	0	3.99
24	3.33	3.15	0	0	0	3.92
25	3.3	0	2.29	0	0	2.83
26	3.3	0	2.29	0	0	2.71
27	14.75	0	1.39	0	0	14.90
28	5.81	0	2.74	0	48.39	6.01
29	5.81	0	2.74	0	48.39	5.95
30	11.25	0	3.98	0	23.44	11.05
31	11.25	0	3.98	0	23.44	10.86
32	8.37	0	1.98	1.55	38.76	8.46
33	8.37	0	1.98	1.55	38.76	8.52
34	4.5	0	2.83	0.83	33.33	5.05
35	4.5	0	2.83	0.83	33.33	4.91
36	16.62	0	0	1.54	7.69	16.34
37	16.62	0	0	1.54	7.69	16.53
38	12.66	0	0	2.34	15.63	12.79
39	12.66	0	0	2.34	15.63	12.95
40	7.63	0	0	3.39	0	8.31
41	7.63	0	0	3.39	0	8.17
42	10.16	0	0	4.03	0	9.73
43	10.16	0	0	4.03	0	9.52
44	9.03	0	1.84	0	0	9.58
45	10.28	0	2.07	0	24.19	11.34
46	8.53	0	3.36	0	35.91	8.5
47	9.81	0	2.98	0.78	31.1	10.39
48	6.44	0	2.41	1.19	36.05	5.93
49	1.75	0	0	0	0	1.86
50	1.75	0	0	0	0	1.84

Table 1

Sample #	Actual Hemolink (g/L)	Hb (g/L)	IL (g/L)	BV (mg/dL)	BR (mg/dL)	Predicted
						Hemolink (g/L)
1	0.85	0	0	0	0	1.03
2	0.85	0	0	0	0	1.03
3	3.41	0	0	0	0	3.73
4	3.41	0	0	0	0	3.56
5	6.52	0	0	0	0	5.92
6	6.52	0	0	0	0	5.89
7	0.87	2.05	0	0	0	1.09
8	0.87	2.05	0	0	0	1.13
9	5.83	0.79	0	0	0	5.54
10	5.83	0.79	0	0	0	5.45
11	2.5	3.94	0	0	0	3.77
12	2.5	3.94	0	0	0	3.50
13	1.31	1.24	0	0	0	1.28
14	1.31	1.24	0	0	0	1.34
15	14.75	0	1.39	0	0	15.24
16	10.56	0	1.42	1.19	20.51	11.36
17	14.64	0	0	1.94	11.66	16.36
18	10.14	0	0	2.87	7.81	10.50
19	8.89	0	0	3.71	0	8.42
20	6.73	0	1.15	2.02	0	5.85
21	1.65	0	1.15	0	0	0.59
22	7.38	0	0.7	0	0	7.02
23	2.9	0	1.37	0	24.19	2.67
24	5.63	0	1.99	0	11.72	4.51
25	4.19	0	0.99	0.78	19.38	4.04
26	2.25	0	1.42	0.42	16.67	1.94
27	8.31	0	0	0.77	3.85	7.78
28	6.33	0	0	1.17	7.81	6.23
29	3.81	0	0	1.69	0	3.58
30	5.08	0	0	2.02	0	4.72

Table 2

Figure 1 is a graphic representation of a linear regression fit of the data generated from the CLHb calibration. The algorithm which was developed for Hb based on this data is as follows:

5 $\text{g/L CLHb} = 23.97(541\text{nm}) - 76.01(558\text{nm}) + 130.84(600\text{nm}) - 113.61(616\text{nm}) + 0.30$

where (X_{nm}) is the first derivative of the absorbance measured at the wavelength specified.

10 Figure 2 provides a graphic representation of the results of linear regression fit for predicted CLHb concentration, for samples not used in the calibration processes.

15 Figure 3 provides a graphic representation of the results of a linear regression fit of the data generated from the true Hb calibration with the presence of cross-linked Hb and other interferents (IL, BR, BV). Table 3 provides the individual data points obtained when performing this calibration. The algorithm which was developed as a result of this calibration is as follows:

20 $\text{g/L Hb} = -0.72 + 30.72(558) - 17.40(570) + 171.14(730)$

where numbers in the parenthesis are the first derivative of the absorbance at the wavelengths (nm) shown.

25 The second part of the present invention is to quantify the relationship between measured amounts of each analyte with respect to the amount of blood substitute present in the serum or plasma specimen, for a particular blood analyzer; these equations which allow one to extrapolate the analyte

CLH:
4.17
7.36
7.36
6.58
6.58
4.21
4.21
0.87
0.87
5.83
5.83
2.50
2.50
1.31
1.31
1.72
1.72
3.33

SUBSTITU

concentration at zero g/L blood substitute must be validated at different levels of the analyte, and if the slopes of the regression lines for different analyte concentration vary with analyte concentration, an average slope value may be used. In this specification, analyte refers to substances routinely measured on a blood analyzer. Tables 4 and 5 give the measured concentrations (all in S. I. units) of various analytes on two different analyzers, in the presence of different levels of CLHb. The analytes which are severely affected by the CLHb are shown in shaded sections of Table 5.

Figures 4 to 7 are graphical representations of linear regression fits, where appropriate, for the measured analytes in response to different amounts of added CLHb. Figure 4 is for total protein (Tprot) when measured on the Kodak Ektachem and the Hitachi 717; Figure 5 is for aspartate aminotransferase (AST) when measured on the Kodak Ektachem and the Hitachi 717; Figure 6 is for alkaline phosphatase (ALP) when measured on the Kodak Ektachem and the Hitachi 717; and Figure 7 is for total bilirubin (Tbili) when measured on the Kodak Ektachem and the Hitachi 717. Where appropriate, the linear regression equations are shown next to the corresponding regression line. To illustrate how this invention works, let us use serum total protein measurement on the Kodak Ektachem as the test which is affected by the presence of CLHb in a patient's serum:

From Figure 4, $y = 2.28x + c$, where

$y = \text{g/L total protein (in presence of CLHb)}$

$x = \text{g/L CLHb present}$

KODAK EKTACHEM								
CLHb (g/L)	Na	K	Cl	HCO3	Ca	Mg	Creat	Urea
0	138	4.3	104	16	2.38	0.65	73	3.9
5	137	4.2	104	16	2.40	0.64	70	3.9
10	137	4.3	104	16	2.42	0.63	68	3.9
15	137	4.2	103	15	2.40	0.62	67	3.9
20	137	4.3	103	15	2.40	0.62	66	3.9
25	136	4.2	103	15	2.41	0.61	66	3.9

HITACHI 717								
CLHb (g/L)	Na	K	Cl	HCO3	Ca	Mg	Creat	Urea
0	140	4.3	105	12	2.09	0.72	86	4.4
5	142	4.3	105	13	2.13	0.75	81	4.5
10	140	4.2	104	14	2.10	0.80	81	4.4
15	139	4.2	103	14	2.10	0.82	82	4.4
20	138	4.2	102	14	2.08	0.83	82	4.5
25	137	4.2	101	14	2.09	0.86	84	4.3

Table 4

KODAK EKTACHEM								
CLHb (g/L)	Glu	TProt	AST	GGT	LDH	CK	ALP	TBili
0	4.6	64	30	45	112	394	68	5
5	4.7	74	24	104	236	404	30	30
10	5.0	82	18	138	219	388	18	51
15	5.2	84	15	78	244	396	17	68
20	5.4	108	13	65	196	388	17	93
25	5.5	121		144			24	119

HITACHI 717								
CLHb (g/L)	Glu	TProt	AST	GGT	LDH	CK	ALP	TBili
0	4.5	64	27	39	100	283	72	2
5	4.9	66	31	36	95	296	61	3
10	5.2	70	37	37	100	299	51	3
15	5.5	73	37	40	100	288	35	4
20	5.7	78	32	40	106	306	29	5
25	6.1	82	33	42	106	316	17	6

Table 5

c = g/L total protein (in presence of zero CLHb)

In this example, c = 62 g/L

The third part of the present invention is combining the first two parts
5 of the invention so as to extrapolate the concentration of the analyte to give
an analyte concentration when there is no blood substitute present. For
illustrative purposes, let us say that the measured amount of CLHb in the
serum specimen is 20 g/L and the measured amount of total protein in the
same specimen is 108 g/L.

10 Question: What is the actual serum total protein?

Answer: From the apparent or measured total protein concentration (in
this case it is 108 g/L), subtract the measured g/L CLHb times
the slope of the regression line (in this case it is 2.28), from the
measured value

15 The answer is 62.4 g/L.

This is in good agreement with the known value of 62 g/L. The accuracy of
this correction will depend on the accuracy of measurement of the blood
substitute, and the reliability of the slope used.

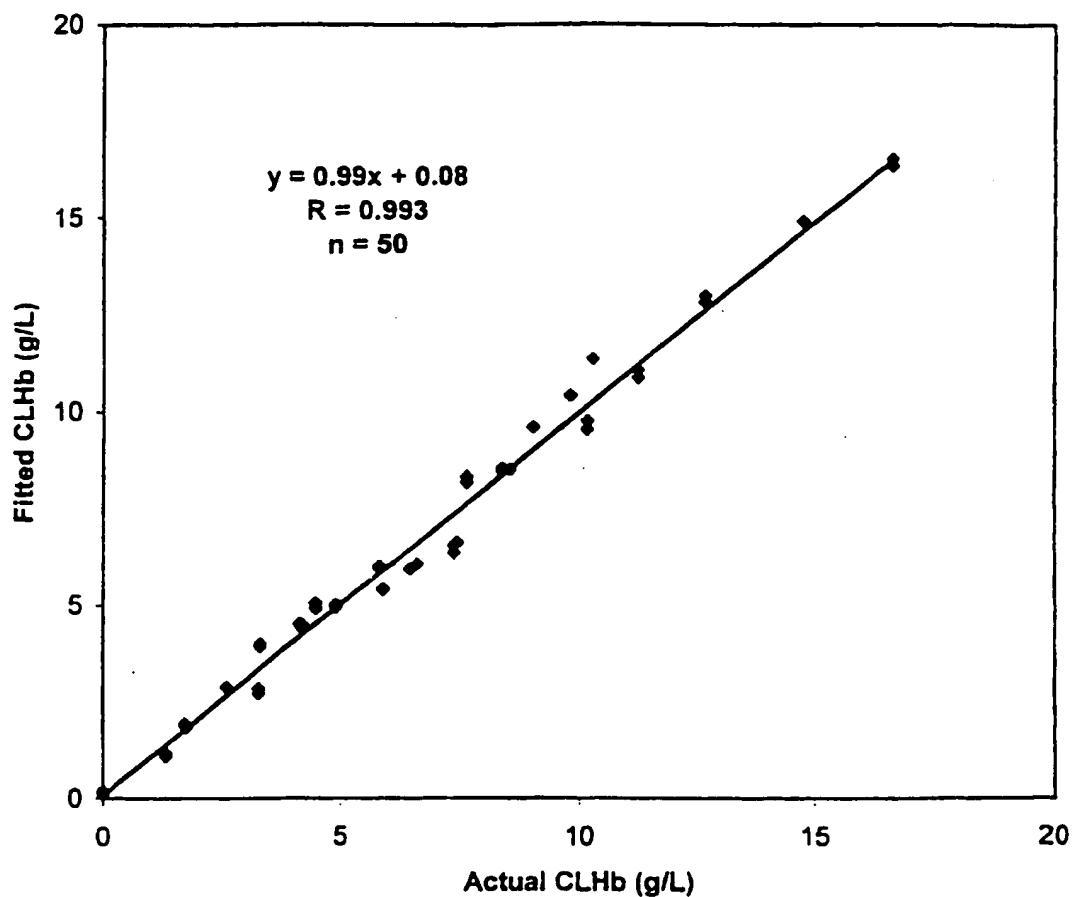
20 As part of this invention, it is also understood that other interferents
can be measured accurately in the presence of blood substitutes. This may
be necessary to determine if other interferents are present in sufficient
quantities that will invalidate correction for blood substitutes. As will be
readily understood by those skilled in the art, several algorithms can be

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developed for each interferent using different groups of wavelengths with the resultant prediction performance by the different algorithms for the same interferent being similar. Also, algorithms can be developed for any interferent or combinations of interferents including blood substitutes, which
5 will enable one to adjust measured analyte concentrations, for the presence of one or more interferents.

While the invention has been particularly shown and described with reference to certain embodiments, it will be understood by those skilled in the art that various other changes in form and detail may be made without departing from the scope of the invention.
10

**Figure 1**

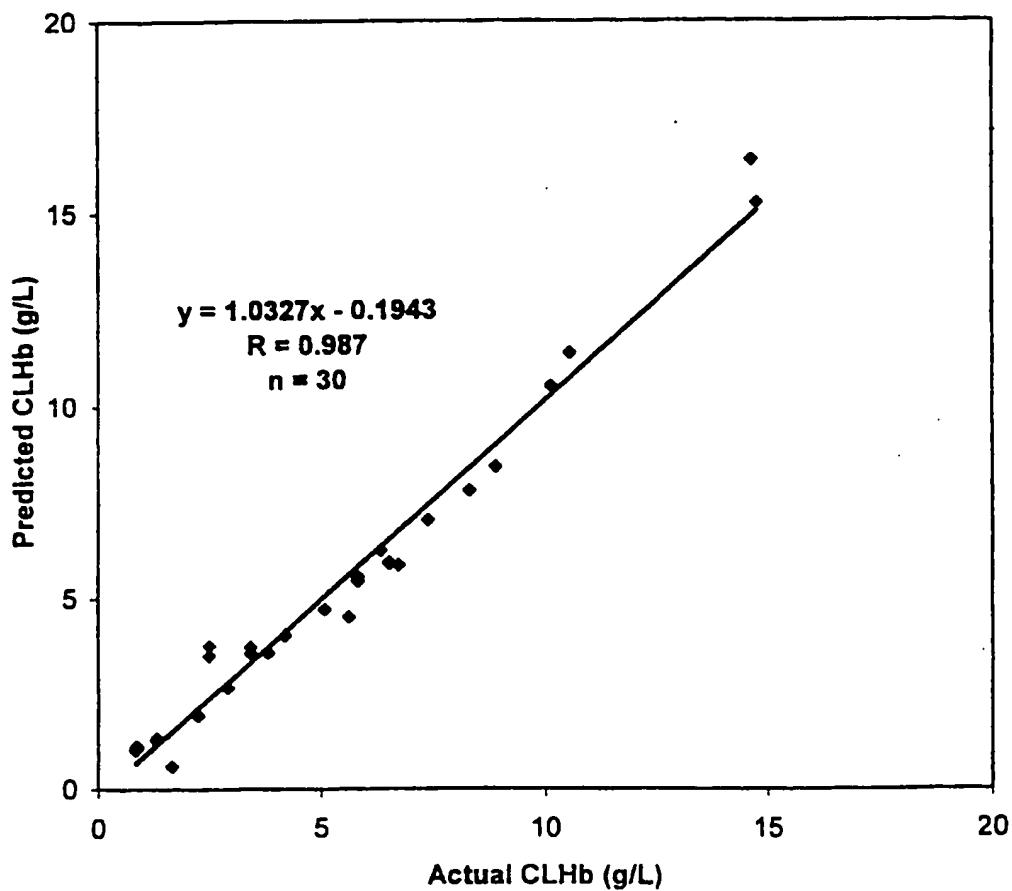


Figure 2

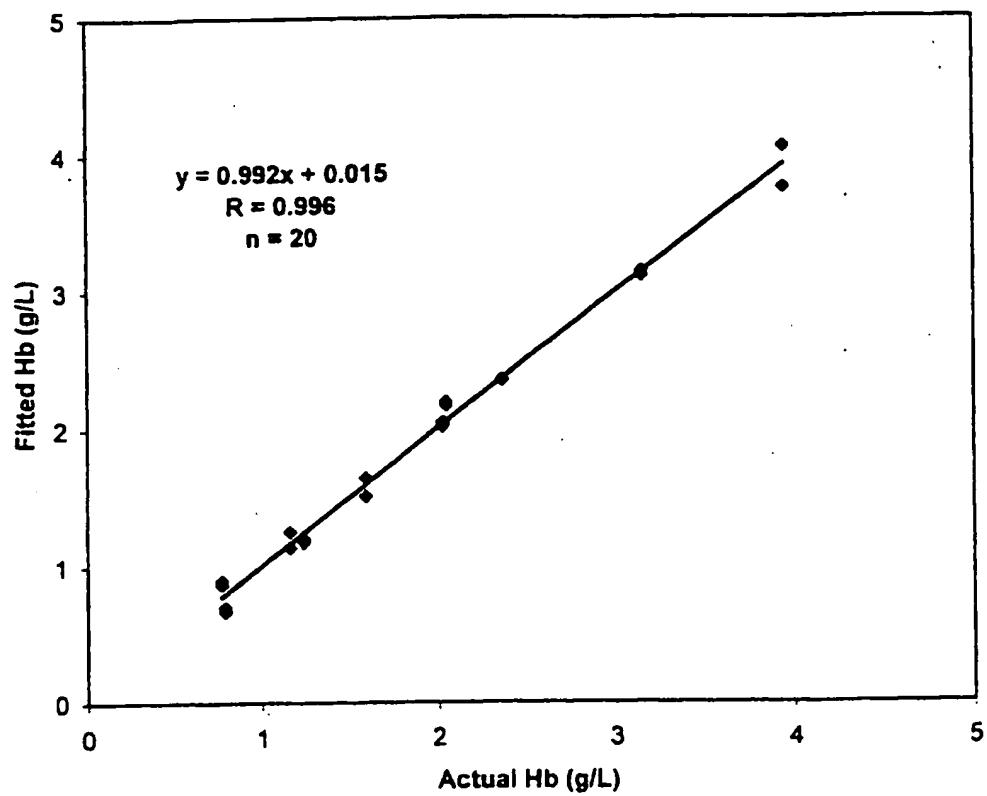
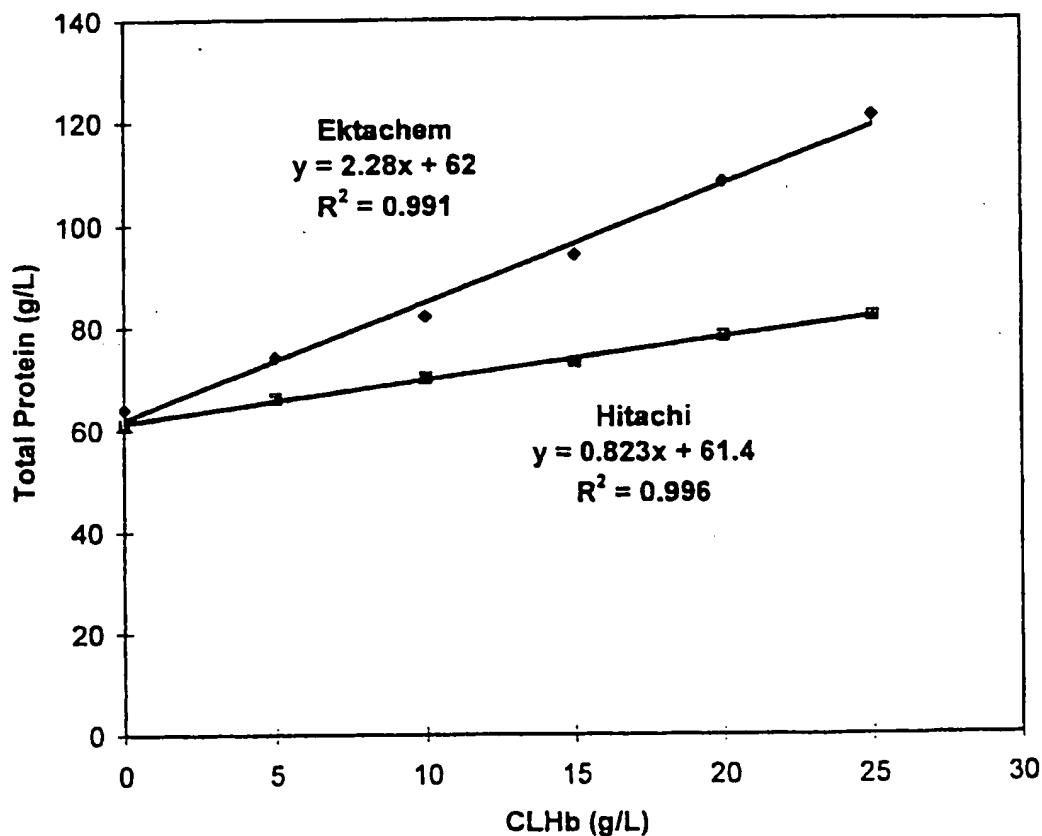
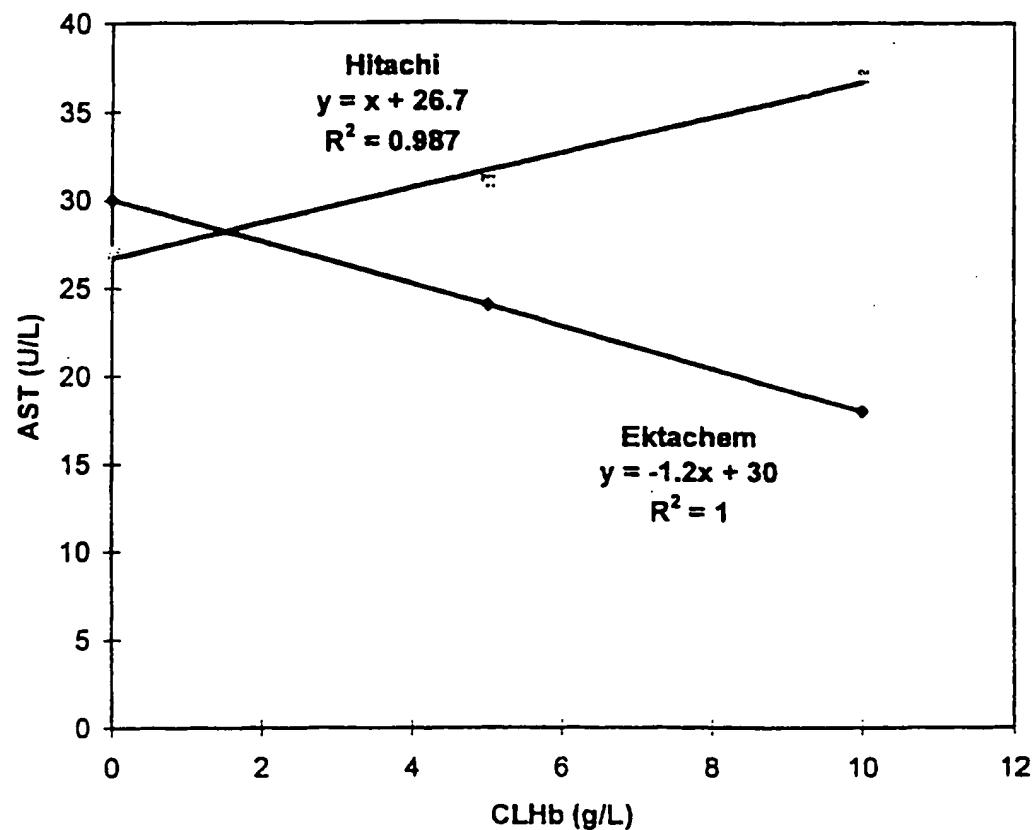


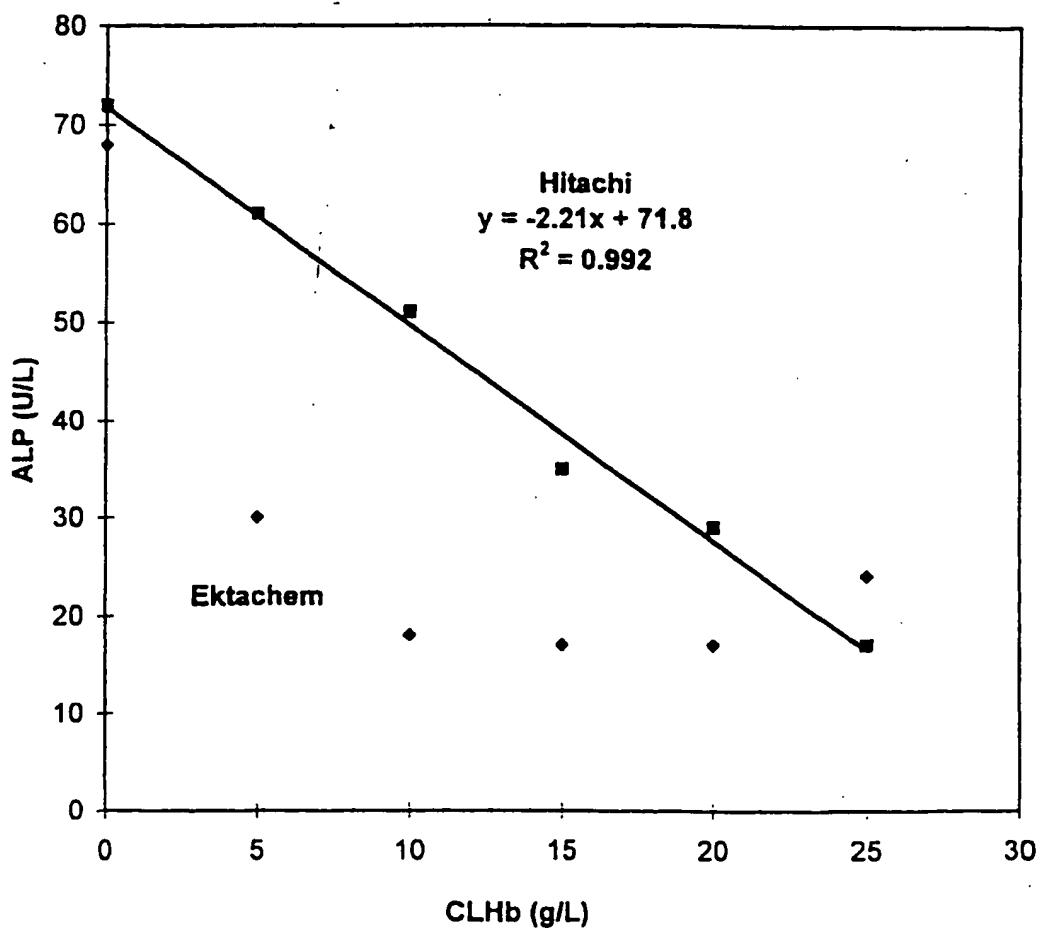
Figure 3

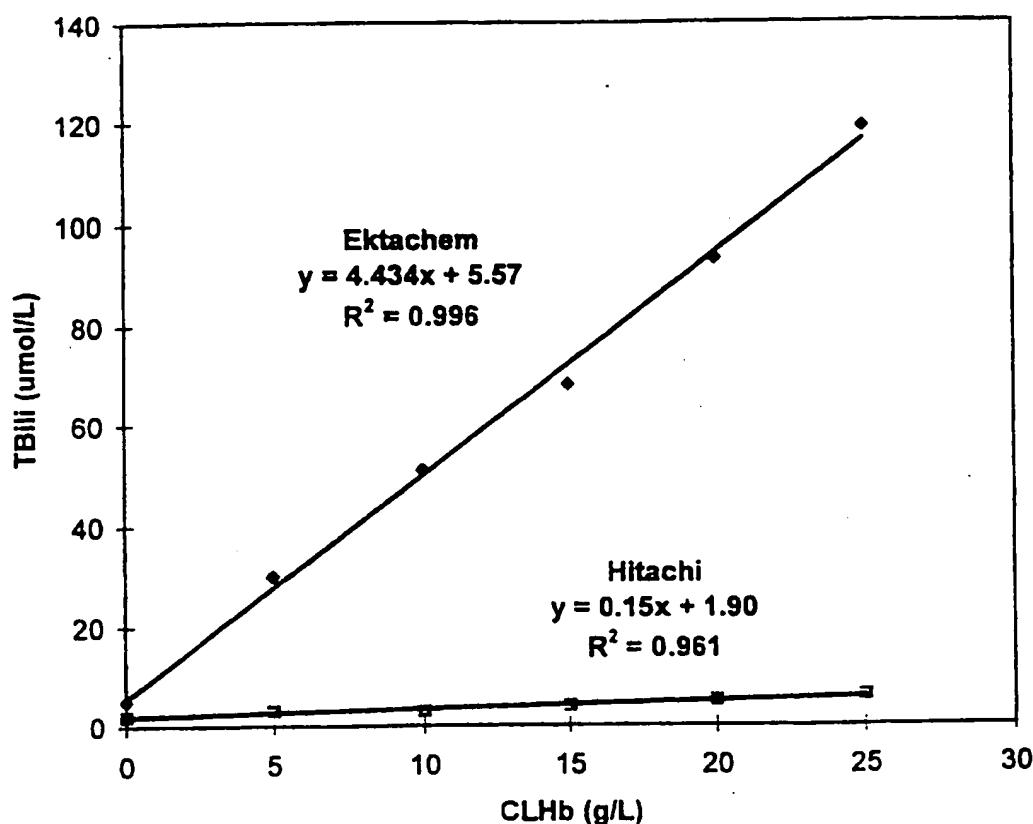
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SUBSTITUTE SHEET (RULE 28)

Effect of CLHb on TProt**Figure 4**

Effect of CLHb on AST**Figure 5**

Effect of CLHb on ALP**Figure 6**

Effect of CLHb on TBili**Figure 7**

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